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## ***Trichoderma* as biocontrol agent - in focus**

### ***Trichoderma* como agente biocontrolador- en foco**

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**ABSTRACT:** *Trichoderma*, a genus of filamentous fungi, is widely used in agriculture due to its various properties and uses, particularly its ability to function as a biocontrol agent against various phytopathogens. The success of *Trichoderma* is based on multiple mechanisms of action, including antibiosis, mycoparasitism, competition for space and nutrients, production of enzymes and secondary metabolites with antimicrobial activity, and stimulation of the plant defense response against pathogens. This mini review focuses on the mechanisms of action of *Trichoderma* as a biocontrol agent and its potential for use in agriculture. Its ability to inhibit the growth of pathogens, stimulate plant growth, and improve soil quality makes it a valuable resource for farmers. Understanding these mechanisms can further improve its use in agriculture and promote sustainable agricultural practices.

**Keywords:** biocontrol, endophyte, mechanism, sustainable.

**RESUMEN:** *Trichoderma*, un género de hongos filamentosos es ampliamente utilizado en la agricultura debido a sus propiedades y usos diversos, destacando su habilidad para funcionar como un agente biocontrolador contra diversos fitopatógenos. El éxito de *Trichoderma* se basa en múltiples mecanismos de acción, que incluyen antibiosis, micoparasitismo, competencia por espacio y nutrientes, producción de enzimas y metabolitos secundarios con actividad antimicrobiana y la estimulación de la respuesta de defensa de las plantas ante los patógenos. Esta mini revisión se centra en los mecanismos de acción de *Trichoderma* como biocontrolador y sus potencialidades para el uso en la agricultura. Este género de hongos filamentosos con capacidad para inhibir el crecimiento de patógenos, estimular el crecimiento de las plantas y mejorar la calidad del suelo lo convierten en un recurso valioso para los agricultores. El conocimiento de estos mecanismos puede ayudar a mejorar aún más su uso en la agricultura y promover prácticas agrícolas sostenibles.

**Palabras clave:** biocontrol, endófito, mecanismo, sostenible.

## 1. INTRODUCTION

Microbial inoculants have emerged as a promising approach for augmenting crop yield and improving the overall quality of agricultural production. *Trichoderma*-based products have captured the attention of researchers and agricultural producers due to their ability to enhance plant resistance to various biotic and abiotic stresses. Additionally, these products can improve the nutritional quality of crops and contribute to sustainable agriculture<sup>(1)</sup>.

Biofungicides, a type of biocontrol agents, have gained attention as a potential solution to reduce the negative impacts, caused by chemical or synthetic fungicides on the environment, animal life, and human health. *Trichoderma* has been widely adopted as a biofungicide by farmers due to its ability to limit the growth of various plant pathogens. For instance, *Trichoderma* has been reported to generate antibiotics and volatile compounds, and to stimulate plant resistance against pathogens. Additionally, it has been shown to compete against other microorganisms in the rhizosphere, while exhibiting mycoparasitic behavior<sup>(2)</sup>.

*Trichoderma* is a dominant active ingredient in more than half of the registered biofungicides that are produced globally, reported approximately 50%, and designed to combat soil-borne pathogens. This reflects the widespread acceptance of *Trichoderma*-based formulations among

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

producers and researchers as a reliable approach for managing plant diseases caused by soil-borne pathogens. The success of these biofungicides is attributed to their effectiveness, safety and compatibility with other crop protection measures<sup>(3)</sup>. *Trichoderma* is one of the most studied antagonists as a biological control agent, as it is capable of controlling a wide range of plant pathogens including bacteria, fungi, insects and nematodes<sup>(4)</sup>.

This fungus is distributed in various ecosystems, where the rhizosphere represents one of the most common ecological niches as it is attracted to certain plant pathogens as prey and to the nutrients exuded by the roots of plants. It can be isolated from all types of agricultural and horticultural fields in different climatic zones due to the diversity of the genus<sup>(2,3,5)</sup>.

In contrast to chemical pesticides, the use of biocontrol agents have a number of advantages, for instance: i) it does not lead to the development of resistance in pathogens; ii) it avoids environmental contamination; iii) it inhibits the proliferation of secondary pests; iv) it is suitable for organic farming practices, and; v) it adheres to regulatory limits regarding maximum chemical residue levels on fruits and vegetables<sup>(6)</sup>.

In view of the role of this fungus in sustainable agriculture, this mini review aims to explore the beneficial aspects of various *Trichoderma* species, highlighting their diverse mechanisms of action.

## **2. MATERIALS AND METHODS**

For the realization of this narrative literature review, databases of national and international journals indexed in Scielo, Latindex 2.0, Scopus, Scimago, and Web of Science were reviewed. Google Scholar and the digital library of the National Council for Science and Technology's Scientific Information Center (CICCO) were utilized. For the searches, keywords such as *Trichoderma*, biocontrol, action mechanisms, mycoparasitism, antibiosis, endoparasitism, defense response induction, biopesticide, bio fungicide, among others, were used.

Subsequently, an Excel spreadsheet was created to organize the data from the articles obtained from various databases. Duplicates were removed, and the articles were thoroughly read. The selection of articles was carried out based on the objectives set forth in this review, scientific articles related to the action mechanisms of *Trichoderma* as a biocontroller and its potential applications in agricultural fields were considered. Works focused on *Trichoderma* as a plant growth promoter were not considered.

The search period ranged from December 2022 to May 2023, and primarily articles from the last 5 years were considered. However, those regarded as worldwide references on the topic and published over 5 years ago were included in this paper. Basic references on the topic outside the specified time period were included.

To obtain information on *Trichoderma*-based products authorized in Paraguay, the website of the regulatory authority was consulted - Servicio Nacional de Calidad y Sanidad Vegetal y de Semillas (SENAVE).

### **3. RESULTS**

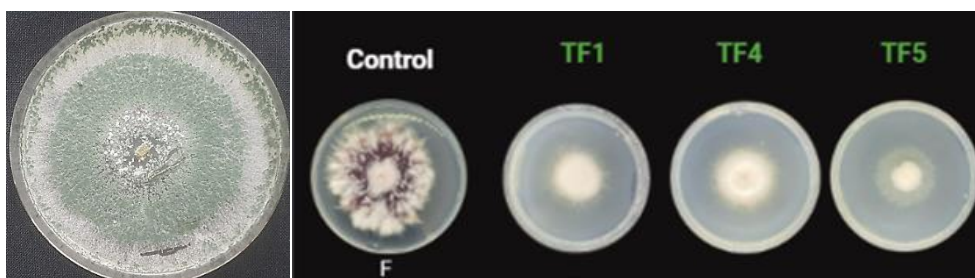
In Google Scholar, using the keywords, a total of 94,500 related documents appeared on the selected topic. Refining the search to the last 5 years reduced the number of documents to 17,400. In the case of the search conducted in Scopus, 327 documents were detected, including 248 articles, 52 literature reviews, 21 book chapters, and 4 conference abstracts. Refining the search to the last 5 years reduced the number of documents to 109. Regarding the search performed in Web of Science, a total of 285 scientific papers were found, including 229 scientific articles, 55 review articles, 2 conference abstracts, 1 book chapter, and 1 rapid communication article. Restricting the search to the last 5 years resulted in 114 scientific articles. After removing duplicates and verifying that the articles were relevant to the objectives established for this review, a total of 81 materials were selected for use in this paper.

### **4. DISCUSSION**

#### **4.1 The Genus *Trichoderma***

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

Persoon published the initial account of the *Trichoderma* genus in 1794<sup>(7)</sup>. As of 2022, 349 *Trichoderma* species had been identified and DNA sequences are available and in public databases like <https://trichokey.com> or and <http://mmit.china-cctc.org><sup>(8)</sup>.



**Imagen 1.** (a) Pure colony of *Trichoderma asperelloides* (b) *Trichoderma asperelloides* as a biocontroller of *Fusarium oxysporum* (own source).

*Trichoderma*, owing to its remarkable adaptability, can be propagated on diverse solid and liquid growth media. The morphological characteristics of the colonies, such as the concentric rings and mycelial pigmentation, are subject to variation depending on the species and culture medium. The rear of *Trichoderma* colonies typically displays hues of colorlessness, beige, yellow, amber, or greenish yellow depending on the growth conditions and species. Thus, these organisms exhibit considerable diversity in their morphological attributes<sup>(9)</sup>.

These fungi reproduce asexually through spores or conidia, which are globular structures, green in color and measure approximately 3-5  $\mu\text{m}$ . their resistance structures are called chlamydo-spores and are usually thick, with a soft green color and intercalary position. Conidiophores have perpendicular or lateral branching in several groups, are green, and measure approximately 62-69 x 3-4.7  $\mu\text{m}$ . The phialides, often in pairs, are long and thin, with up to four terminal verticils of phialides<sup>(9,10)</sup>.

*Trichoderma* produces abundant conidia to maintain long periods of vegetative growth. However, the transition from mycelium to conidium is determined by a combination of factors capable of triggering this change. Nutritional environment, pH, metabolite production, light, and even the fungus' own metabolism, are some of the factors responsible for the way conidiation occurs. This process is critical for the survival of the fungus, yet it has been shown that the conidial response varies widely, depending on the metabolic adaptation of each species to the environment<sup>(11)</sup>.

Comparative genetic studies have demonstrated that *Trichoderma* has undergone genome remodeling to enhance its ability to rapidly colonize and successfully compete in new habitats. As such, it may be misleading to associate specific biological activities, such as metabolite production, with species that have been described under outdated names within the current taxonomic classification of *Trichoderma*<sup>(12)</sup>.



## 4.2 Mechanisms of action and biological control agent

It is important to note that the efficacy-based biocontrol agents can be improved through the selection of competent strains and optimal formulations of organisms adapted to various agroecosystems. However, it is crucial to enhance the performance levels of this antagonist and acknowledge that the outcomes of biological control rely on the interaction between the pathogen and *Trichoderma*. Additionally, it should be considered that *Trichoderma* is most effective in controlling pathogens preventively<sup>(6)</sup>.

According to the comparative analysis of the genomes of three *Trichoderma* species (*T. virens*, *T. reesei*, and *T. atroviride*) commonly utilized as biocontrol agents in agriculture, it has been revealed that the original lifestyle of the fungal genus was mycoparasitism<sup>(4)</sup>. The fungi produce various metabolites such as antibiotics, mycotoxins, and phytotoxins, which aid in its antagonistic effects. Additionally, the fungus releases enzymes such as glucanases, chitobioses, and chitinases, as well as antibiotics like viridin, gliotoxin, or peptaibols<sup>(13,14)</sup>

Mechanism of action	Description	Pathogen specificity	References
Mycoparasitism	<p>The hyperparasite is dependent on the host fungus and acquires nutrients through haustoria without inducing host cell death. These attacks are initiated by the <i>Trichoderma</i> fungi through the penetration of the cell wall of the pathogen, followed by the subsequent degradation of its cell components. <i>Trichoderma</i> have conventionally been regarded as necrotrophic mycoparasites, and research has mainly concentrated on the degradation of the host's cell wall. Nonetheless, a mode of action analogous to "hemibiotrophy" has been proposed, and evidence suggests that the fungal cell wall of the host is not extensively harmed during interactions with</p>	Pathogen specific interaction	(14-18).

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

	<i>Trichoderma</i> . Conversely, digestion and mobilization of the cellular contents appear to be critical.		
Antibiosis	Production of low-molecular-weight volatile or non-volatile antibiotics or diffusible compounds. These substances that inhibit or reduce the growth and/or proliferation of phytopathogens. More than 90 metabolites have been reported in <i>Trichoderma</i> species including Trichorzianin TA, Trichorzianin TB, 6-pentyl-2H-pyran-2-one, Trichodermin, Cyclonerodiol, Pachybasin and others.	Broad	(2,14,16,17,19–21).
Competition	<i>Trichoderma</i> compete with phytopathogens for space and essential nutrients. It is capable of colonizing the root rhizosphere and effectively competing with other microorganisms for nutrients secreted by the soil. In addition, it	Broad	(14,16,22–24).

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

	has the ability to promote plant growth. <i>Trichoderma</i> spp. releases siderophores that sequester Fe <sup>3+</sup> , making them inaccessible to pathogens.		
Induction of plant defenses and endophytism	Activation of host defense mechanisms against diseases and other stresses. <i>Trichoderma</i> can interact with plants and elicit a defense response against pathogens or disease. The use of <i>Trichoderma</i> results in the production and accumulation of enzymes, secondary metabolites, and signaling molecules, such as salicylic acid (SA), ethylene (ET), and jasmonic acid (JA), leading to enzymatic and morphological changes within the host plant. Ultimately, this results in the induction of induced systemic resistance in the plant. The interaction between <i>Trichoderma</i> and the plant is dependent on	Specific to broad	(14–17,27–29).

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

	<p>various factors, including the strain, its concentration, the plant material, developmental stage, and the timing of interaction. <i>Trichoderma</i> contains genes that are expressed in plants to help them manage diseases and impart resistance to environmental stressors. Commonly used marker genes in this defense include PDF1.2 (Plant defensin 1.2), Thi2.1 (Thionin), or Chib (Chitinase B). The salicylic acid-mediated systemic acquired resistance (SAR) results in the expression of pathogenesis-related genes (PR).</p>		
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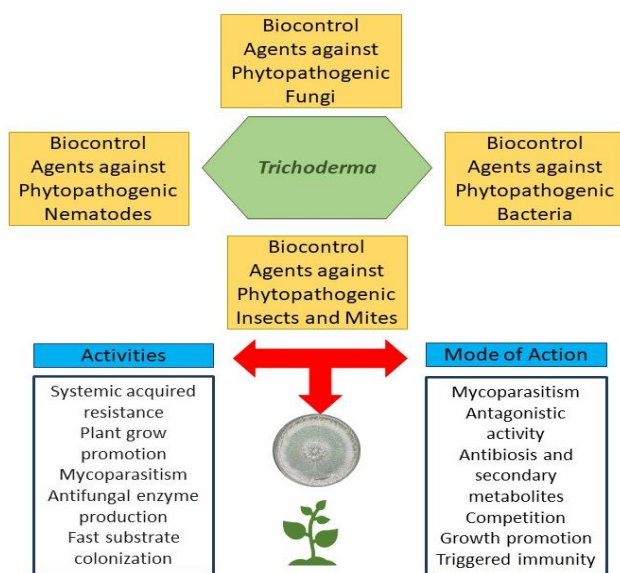
**Table 1.** Mechanism of action of *Trichoderma* as a biocontrol organism.

*Trichoderma* employs various intricate direct and indirect mechanisms of biocontrol to combat biotic stresses posed by a broad range of pathogenic microorganisms, including fungi, bacteria, insects, and nematodes, as well as abiotic stresses resulting from unfavorable environmental conditions<sup>(30)</sup>. Not all *Trichoderma* species possess the capacity to modulate plant growth

and physiology due to the wide range of symbiotic interactions between microorganisms and plants. Additionally, the response of the antagonist in terms of the production of secondary metabolites may either promote or inhibit plant growth, thus contributing to the complexity of this relationship<sup>(31)</sup>.

### 4.3 Potential biocontrol of *Trichoderma* in agriculture

*Trichoderma* has become a key ally in the integrated management of pests in crops, thanks to its diverse mechanisms for controlling without negatively impacting the environment or generating resistance in pests<sup>(32)</sup>.



**Figure 1.** Roles of *Trichoderma* in sustainable plant disease management.

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

*Trichoderma* has been reported as an effective agent for controlling phytopathogenic fungi for several years<sup>(6)</sup>; Its biocontrol effect has been mentioned against *Rhizoctonia solani*<sup>(33)</sup>; *Fusarium graminearum*<sup>(34,35)</sup>; *Pythium*<sup>(36–38)</sup>; *Fusarium oxysporum*<sup>(4,39,40)</sup>; *Botrytis cinerea*<sup>(41–43)</sup>; *Sclerotium* and *Macrophomina*<sup>(44,45)</sup>; *Colletotrichum*<sup>(46–48)</sup>; *Sclerotinia*<sup>(49–51)</sup> and others.

Regarding its effect in the control of phytopathogenic bacteria, *Trichoderma* has been mentioned as effective for *Ralstonia solanacearum*<sup>(52,53)</sup>; *Xanthomonas*<sup>(54,55)</sup>, however, the literature referring to the control of bacteria with *Trichoderma* is scarce in comparison with the applications for the control of phytopathogenic fungi.

*Trichoderma* has been mentioned as a biocontrol agent for species of nematodes of the genus *Meloidogyne* including *Meloidogyne incognita*<sup>(56–58)</sup> and others like *Heterodera*<sup>(59)</sup>.

Regarding the use of *Trichoderma* as an entomopathogen, its efficacy against *Leucinodes orbonalis*<sup>(60)</sup>, Regarding the use of *Trichoderma* as an entomopathogen, its efficacy against *Leucinodes orbonalis*<sup>(61)</sup>, *Spodoptera littoralis* and *Macrosiphum euphorbiae*<sup>(62)</sup>. Other authors have mentioned in reviews on the subject the control of mites, hemiptera, coleoptera, diptera, orthoptera and lepidoptera, among others<sup>(4)</sup>.

## Interaction of *Trichoderma* with other beneficial organisms

*Trichoderma* is a well-known and extensively studied genus of fungi that plays a significant role in biological control strategies for managing plant diseases. Its interactions with other biocontrol organisms are of particular interest due to their potential synergistic effects on enhancing plant health and protection<sup>(30,32,63)</sup>.

This biocontroller, exhibits various modes of interaction with other biocontrol agents, such as beneficial bacteria and mycorrhizal fungi. These interactions can lead to complementary or cooperative activities that bolster the overall effectiveness of integrated pest management strategies<sup>(4,64,65)</sup>.

Some key aspects of *Trichoderma's* interactions with other biocontrol organisms include:

**Antagonistic Interactions:** *Trichoderma* is often recognized for its mycoparasitic capabilities, meaning it can attack and inhibit the growth of pathogenic fungi through mechanisms such as competition for resources, secretion of antifungal compounds, and direct parasitism. When combined with other biocontrol agents that possess different modes of action, the collective antagonistic effect can target a broader range of pathogens and contribute to more robust disease suppression<sup>(7,30,32,63,66)</sup>.



**Synergistic Effects:** Certain strains of *Trichoderma* have been found to enhance the performance of other biocontrol organisms. For example, when used in conjunction with beneficial bacteria, *Trichoderma* can promote the colonization of these bacteria on plant surfaces, thereby increasing the potential for disease prevention. This synergy can lead to improved establishment and persistence of biocontrol populations<sup>(63,64)</sup>.

**Plant Growth Promotion:** *Trichoderma's* ability to enhance plant growth and nutrient uptake can also complement the activities of mycorrhizal fungi. Mycorrhizal fungi form symbiotic relationships with plant roots, aiding in nutrient acquisition. *Trichoderma* can improve root system development, making plants more receptive to mycorrhizal colonization and leading to greater overall plant health<sup>(67,68)</sup>.

**Induced Systemic Resistance (ISR):** *Trichoderma's* interactions with other biocontrol agents can trigger systemic defense responses in plants, a phenomenon known as induced systemic resistance. This mechanism enhances the plant's innate ability to ward off pathogens, making it more resilient to disease attacks. When integrated with other biocontrol agents, this systemic defense response can create a multi-layered defense strategy for plants<sup>(30,66,69,70)</sup>.

**Environmental Adaptation:** Different biocontrol organisms, including *Trichoderma* strains, possess varying environmental tolerances and

preferences. In some cases, combining strains adapted to different conditions can result in broader and more reliable disease management across diverse environmental settings<sup>(71-75)</sup>.

In Paraguay, a variety of products based on *Trichoderma* are commercially available and described in Table 2, which lists products legally registered by the Servicio Nacional de Calidad y Sanidad Vegetal (SENAVE). These products are recommended for different crops and pathogens, containing various species of *Trichoderma* as active ingredients. They are primarily marketed for the control of phytopathogenic fungi and nematodes<sup>(76)</sup>

Product and Ref.	<i>Trichoderma</i> specie	Formulation	Use class	Toxicology	Manufacturer	Origen	Pathogens	Crops	Ref.
ECOTRICH	<i>T. harzianum</i>	WP Wettable Powder	Biological fungicide	IV	Ballagro Agro Tecnología Ltda	Brazil	<i>Sclerotinia sclerotiorum</i> , <i>Macrophomina phaseolina</i>	Soybean, alfalfa	(76,77)
TRICHODERMIL WP 1306	<i>T. harzianum</i>	WP Wettable Powder	Biological fungicide and nematicide	III	Koppert Do Brasil Holding Ltd.	Brazil	<i>Rhizoctonia solani</i> , <i>Fusarium solani</i> , <i>Sclerotinia sclerotioum</i> , <i>Thielaviopsis paradoxa</i> , <i>Meloidogyne</i> sp. and <i>Pratylenchus</i> sp.	Extensive and intensive crops	(76,78)
STIMUCONTROL	<i>T. harzianum</i>	CS Concentrated Suspension	Biological fungicide	III	Simbiose Industria E Comercio De Fertilizantes E Insumos Microbiológicos Ltda	Brazil	<i>Rhizoctonia solani</i> , <i>Sclerotinia clerotiorum</i>	Extensive and intensive crops	(76,79)
RIZODERMA	<i>T. harzianum</i>	AL Liquid	Fungicide	IV	Rizobacter Argentina S.A.	Argentina	<i>Cercosporakikuchii</i> <i>Phomopsis</i> <i>Fusarium</i> spp. <i>Fusarium</i> spp. <i>Alternaria</i> spp. <i>Bipolaris</i> spp. <i>Fusarium graminearum</i> <i>Drechsleratriticirepentis</i> <i>Bipolarissorokiniana</i> <i>Tilletialaevivis</i> <i>Ustilagotritici</i>	Soybean, rice, wheat	(76,80)

Rev. Soc. cient. Parag. Jun. 2024;29(1):136-174

LALSTOP QUALITY WG	<i>T. asperellun</i>	WP Wettable Powder	Microbiologi cal fungicide	III	LallemandSoluções Agrobiológicas Ltda.	Brazil	<i>Botrytis cinérea,</i> <i>Didymellabryoniae,</i> <i>Pythium, Rhizoctonia,</i> <i>Phytophthora and</i> <i>Fusarium, Verticillium,</i> <i>Macrophominaphaseolina</i>	Large variety of fruit and vegetable crops such as tomato, pepper, cucumber, lettuce, herbs and ornamentals, cucurbit crops such as cucumber and melons, herb and ornamental major fruit, strawberry and sweet potato.	(76,81).
BIO-R1	<i>T. asperellun</i>	EC EmulsifiableConc- entrate	Microbiological fungicide	IV	Vittia Fertilizantes E Biológicos S.A.	Brazil	No data	No data	(76).
HULKGREEN	<i>T. harzianum</i>	CS Concentrated Suspension	Biological fungicide	IV	Agro Advance Technology S.A.	Argentina	<i>Fusarium sp.,</i> <i>Sclerotinia sp.</i>	Extensive and intensive crops	(76,82).

Rev. Soc. cient. Parag. Jun. 2024;29(1):136-174

RIZODERMA MAX	<i>T. harzianum</i>	AL Liquid	Therapic fungicide for seed treatment	IV	Rizobacter Argentina S.A.	Argentina	<i>Cercosporakichii</i> <i>Phomopsis</i> <i>Fusarium spp.</i> <i>Fusarium spp.</i> <i>Alternaria spp.</i> <i>Bipolaris spp.</i> <i>Fusarium graminearum</i> <i>Drechsleratritiicrepentis</i> <i>Bipolarissorokiniana</i> <i>Tilletia laevis</i> <i>Ustilago tritici</i>	Soybean, rice, wheat	(76,80)
BIO-FORCE	<i>T. asperellum</i>	EC EmulsifiableConc- entrate	Nematicide, microbiologi cal fungicide	IV	Vittia Fertilizantes E Biológicos S.A.	Brazil	No data	No data	(76)
TRICHOMBAT	<i>T. atroviride</i>	WP Wettable Powder	Microbiologi cal fungicide	IV	Innova Ltda	Brazil	No data	No data	(76)
AQ® TRC	<i>T. capillare</i>	AL Liquid	Biofungicide	Not applicable	Aquafree S.R.L	Paragua y	<i>Phytophthora sp.</i> , <i>Rhizoctonia sp.</i> , <i>Sclerotium sp.</i> , <i>Fusarium sp.</i> , <i>Rosellinia sp.</i> , <i>Botrytis sp.</i> , <i>Alternaria sp.</i> , <i>Cercosporasp.</i> , <i>Colletotrichum sp.</i> , <i>Peronospora sp.</i> , <i>Oidium sp.</i> , <i>Pyricularia sp.</i>	Soybean, rice, corn, sorghum, rice, pastures, cotton, sugar cane, sesame, chia, yerba mate, bananas, perennial	(76,83)

								plants, horticultur al and ornamental crops	
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**Table 2.** *Trichoderma* based commercial bioformulations registered in Paraguay

## 5. CONCLUSION

The potential of *Trichoderma* species to foster sustainable agriculture is of paramount importance. These fungi exhibit a myriad of mechanisms that can profoundly influence soil health, plant development, and the ability to ward off diseases. *Trichoderma's* multifaceted role as a biocontrol agent, enhancer of nutrient assimilation, and fortifier against stress holds great promise for ushering in sustainable agricultural paradigms.

Moreover, the application of *Trichoderma* as a biofertilizer offers a pivotal advantage by diminishing reliance on synthetic fertilizers notorious for their environmentally detrimental impacts. This collective prowess positions *Trichoderma* as a compelling and environmentally conscious substitute for conventional agricultural methodologies.

Furthermore, the wealth of scientific articles available online underscores the extensive body of research dedicated to unraveling the potentialities of *Trichoderma*. These articles expound on the intricate ways these fungi can augment soil microbial communities, amplify plant growth, and bolster disease resistance. This robust scientific foundation not only validates *Trichoderma's* efficacy but also highlights the ongoing commitment to harnessing its capabilities for sustainable agricultural advancement.

The dynamic attributes of *Trichoderma* species underscore their potential as a cornerstone of sustainable agricultural practices. Their capacity to revolutionize disease control, amplify nutrient efficiency, and alleviate ecological strain paves a progressive path toward a greener and more sustainable future in agriculture.

## **6. FINANCIAL DISCLOSURE**

The present research was carried out with the company's own funding.

## **7. DECLARATION OF CONFLICTS OF INTEREST**

The authors declare that they have no conflicts of interest.

## **8. STATEMENT OF AUTHORS**

The authors approve the final version of the article.

## **9. EDITOR RESPONSABLE**

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## **10. BIBLIOGRAPHIC REFERENCES**

1. Fiorentino N, Ventorino V, Woo SL, Pepe O, De Rosa A, Gioia L, et al. *Trichoderma*-based biostimulants modulate rhizosphere microbial populations and improve N uptake efficiency, yield, and nutritional quality of leafy vegetables. *Front Plant Sci* [Internet]. 2018;9(June):1–15. Available from:

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco



<https://www.frontiersin.org/articles/10.3389/fpls.2018.00743/full>

2. Kubheka BP, Ziena LW, Kubheka BP, Ziena LW. *Trichoderma*: A Biofertilizer and a Bio-Fungicide for Sustainable Crop Production. *Trichoderma - Technol Uses* [Internet]. 2022 Feb 24 [cited 2023 Apr 10]; Available from: <https://www.intechopen.com/chapters/80375>
3. Shahriar SA, Islam MN, Chun CNW, Kaur P, Rahim MA, Islam MM, et al. Microbial Metabolomics Interaction and Ecological Challenges of *Trichoderma* Species as Biocontrol Inoculant in Crop Rhizosphere. *Agronomy* [Internet]. 2022;12(4):1–17. Available from: <https://www.mdpi.com/2073-4395/12/4/900>
4. Poveda J. *Trichoderma* as biocontrol agent against pests: New uses for a mycoparasite. *Biol Control* [Internet]. 2021 Aug 1;159:104634. Available from: <https://doi.org/10.1016/j.biocontrol.2021.104634>
5. Nafaa M, Rizk SM, Aly TAGA, Rashed MAS, Abd El-Moneim D, Ben Bacha A, et al. Screening and Identification of the Rhizosphere Fungal Communities Associated with Land Reclamation in Egypt. *Agric* [Internet]. 2023;13(1). Available from: <https://www.mdpi.com/2077-0472/13/1/215>
6. Ferreira F V., Musumeci MA. *Trichoderma* as biological control agent: scope and prospects to improve efficacy. *World J Microbiol Biotechnol* [Internet]. 2021;37(5):1–17. Doi: <https://doi.org/10.1007/s11274-021-03058-7>
7. Mukhopadhyay R, Kumar D. *Trichoderma*: a beneficial antifungal agent and insights into its mechanism of biocontrol potential. *Egypt J Biol Pest Control* [Internet]. 2020 Dec 1;30(1):1–8. Available from: <https://ejbpc.springeropen.com/articles/10.1186/s41938-020-00333-x>
8. Guzmán-Guzmán P, Kumar A, de los Santos-Villalobos S, Parra-Cota FI, Orozco-Mosqueda M del C, Fadiji AE, et al. *Trichoderma* Species: Our Best Fungal Allies in the Biocontrol of Plant Diseases—A Review. *Plants* 2023, Vol 12, Page 432 [Internet]. 2023 Jan 17;12(3):432. Available from: <https://www.mdpi.com/2223-7747/12/3/432/htm>
9. Siddiquee S. Recent Advancements on the Role and Analysis of Volatile Compounds (VOCs) from *Trichoderma*. *Biotechnol Biol Trichoderma*. 2014 Jan

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

1;139–75.

10. Bissett J. A revision of the genus *Trichoderma* . II. Infrageneric classification. *Can J Bot* [Internet]. 1991;69(11):2357–72. Available from: <https://cdnsiencepub.com/doi/10.1139/b91-297>
11. Steyaert JM, Weld RJ, Mendoza-Mendoza A, Stewart A. Reproduction without sex: Conidiation in the filamentous fungus *Trichoderma*. *Microbiology* [Internet]. 2010;156(10):2887–900. Available from: <https://pubmed.ncbi.nlm.nih.gov/20688823/>
12. Woo SL, Hermosa R, Lorito M, Monte E. *Trichoderma*: a multipurpose, plant-beneficial microorganism for eco-sustainable agriculture. *Nat Rev Microbiol* [Internet]. 2023;21(5):312–26. Available from: <https://www.nature.com/articles/s41579-022-00819-5>
13. Dou K, Lu Z, Wu Q, Ni M, Yu C, Wang M, et al. MIST: A multilocus identification system for *Trichoderma*. *Appl Environ Microbiol* [Internet]. 2020;86(18):1–13. Available from: <https://pubmed.ncbi.nlm.nih.gov/32680870/>
14. Manzar N, Kashyap AS, Goutam RS, Rajawat MVS, Sharma PK, Sharma SK, et al. *Trichoderma*: Advent of Versatile Biocontrol Agent, Its Secrets and Insights into Mechanism of Biocontrol Potential. *Sustain* [Internet]. 2022;14(19):1–32. Available from: <https://www.mdpi.com/2071-1050/14/19/12786>
15. Harman GE, Herrera-Estrella AH, Horwitz BA, Lorito M. Special issue: *Trichoderma*--from basic Biology to Biotechnology. *Microbiology* [Internet]. 2012 Jan;158(Pt 1):1–2. Available from: <https://pubmed.ncbi.nlm.nih.gov/22210803/>
16. Olowe OM, Nicola L, Asemoloye MD, Akanmu AO, Babalola OO. *Trichoderma*: Potential bio-resource for the management of tomato root rot diseases in Africa. *Microbiol Res* [Internet]. 2022;257(February):126978. Available from: <https://doi.org/10.1016/j.micres.2022.126978>
17. Köhl J, Kolnaar R, Ravensberg WJ. Mode of action of microbial biological control agents against plant diseases: Relevance beyond efficacy. *Front Plant Sci* [Internet]. 2019;10(July):1–19. Available from: <https://www.frontiersin.org/articles/10.3389/fpls.2019.00845/full>

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

18. Mukherjee PK, Nautiyal CS, Mukhopadhyay AN. Molecular Mechanisms of Biocontrol by *Trichoderma* spp. 2008;243–62. Available from: [https://link.springer.com/chapter/10.1007/978-3-540-75575-3\\_10](https://link.springer.com/chapter/10.1007/978-3-540-75575-3_10)
19. Elena Cardoza R, Hermosa R, Antonio Vizcaino J, Sanz L, Cardoza R-E, Hermosa M-R, et al. Secondary metabolites produced by *Trichoderma* and their importance in the biocontrol process. *Microorg Ind Enzym biocontrol* [Internet]. 2005;1(January):1–22. Available from: <https://www.researchgate.net/publication/284802240>
20. Aro N, Ilmén M, Saloheimo A, Penttilä M. ACEI of *Trichoderma reesei* Is a Repressor of Cellulase and Xylanase Expression. *Appl Environ Microbiol* [Internet]. 2003 Jan 1;69(1):56. Available from: [/pmc/articles/PMC152388/](https://pubmed.ncbi.nlm.nih.gov/152388/)
21. Garnica-Vergara A, Barrera-Ortiz S, Muñoz-Parra E, Raya-González J, Méndez-Bravo A, Macías-Rodríguez L, et al. The volatile 6-pentyl-2H-pyran-2-one from *Trichoderma atroviride* regulates *Arabidopsis thaliana* root morphogenesis via auxin signaling and ETHYLENE INSENSITIVE 2 functioning. *New Phytol* [Internet]. 2016;209(4):1496–512. Available from: <https://pubmed.ncbi.nlm.nih.gov/26568541/>
22. Hemming BC. Microbial-iron interactions in the plant rhizosphere. an overview. *J Plant Nutr* [Internet]. 1986;9(3–7):505–21. Doi: <https://www.tandfonline.com/doi/abs/10.1080/01904168609363462>
23. J B Neilands and, Leong SA. Siderophores in Relation to Plant Growth and Disease. <https://doi.org/10.1146/annurev.pp37060186001155> [Internet]. 2003 Nov 28;37(1):187–208. Doi: <https://www.annualreviews.org/doi/abs/10.1146/annurev.pp.37.060186.001155>
24. Leong SA. Siderophores in relation to plant growth and disease. 1986;(35):187–208. Available from: <https://www.annualreviews.org/doi/abs/10.1146/annurev.pp.37.060186.001155>
25. Monfil, V.O. and Casas-Flores S. Molecular Mechanisms of Biocontrol in *Trichoderma* spp. and Their Applications in Agriculture, In Gupta, M.S.V., Herrera-Estrella, A., Upadhyay, R., Druzhinina, I. and Tuohy, M., Eds., *Biotechnology and Biology of Trichoderma*, Elsevier, Amsterdam, [Internet].

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

2014. Available from:

[https://www.scirp.org/\(S\(i43dyn45teexjx455qlt3d2q\)\)/reference/referencespapers.aspx?referenceid=2330209](https://www.scirp.org/(S(i43dyn45teexjx455qlt3d2q))/reference/referencespapers.aspx?referenceid=2330209)

26. Hernandez-Flores JL, Melo JGB, Hernández AC, López MAR, Gutiérrez CS, Gomez SR, et al. Isolation and Characterization of Mercury Resistant Trichoderma Strains from Soil with High Levels of Mercury and Its Effects on *Arabidopsis thaliana* Mercury Uptake. *Adv Microbiol.* 2018;08(07):600–13.
27. Atanasova L, Crom S Le, Gruber S, Coulpier F, Seidl-Seiboth V, Kubicek CP, et al. Comparative transcriptomics reveals different strategies of Trichoderma mycoparasitism. *BMC Genomics* [Internet]. 2013;14(1). Available from: <https://bmcgenomics.biomedcentral.com/articles/10.1186/1471-2164-14-121>
28. Bolton MD, Thomma BPHJ, Nelson BD. Sclerotinia sclerotiorum (Lib.) de Bary: Biology and molecular traits of a cosmopolitan pathogen. *Mol Plant Pathol* [Internet]. 2006;7(1):1–16. Available from: <https://pubmed.ncbi.nlm.nih.gov/20507424/>
29. Ding Z, Wang X, Kong FD, Huang HM, Zhao YN, Liu M, et al. Overexpression of Global Regulator Talae1 Leads to the Discovery of New Antifungal Polyketides From Endophytic Fungus Trichoderma afroharzianum. *Front Microbiol.* 2020 Dec 23;11:3351.
30. Tyśkiewicz R, Nowak A, Ozimek E, Jaroszuć-Ścisiel J. Trichoderma: The Current Status of Its Application in Agriculture for the Biocontrol of Fungal Phytopathogens and Stimulation of Plant Growth. *Int J Mol Sci* [Internet]. 2022;23(4). Available from: <https://www.mdpi.com/1422-0067/23/4/2329>
31. Alfiky A, Weisskopf L. Deciphering trichoderma–plant–pathogen interactions for better development of biocontrol applications. *J Fungi* [Internet]. 2021;7(1):1–18. Available from: <https://pubmed.ncbi.nlm.nih.gov/33477406/>
32. Zin NA, Badaluddin NA. Biological functions of Trichoderma spp. for agriculture applications. *Ann Agric Sci* [Internet]. 2020;65(2):168–78. Doi: <https://doi.org/10.1016/j.aos.2020.09.003>
33. Naeimi S, Okhovvat SM, Javan-Nikkhah M, Vágvölgyi C, Khosravi V, Kredics L.

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

Biological control of *Rhizoctonia solani* AG1-1A, the causal agent of rice sheath blight with *Trichoderma* strains. *Phytopathol Mediterr* [Internet]. 2010;49(3):287–300. Available from:

<https://oajournals.fupress.net/index.php/pm/article/view/5372>

34. He A le, Liu J, Wang X hua, Zhang Q guo, Song W, Chen J. Soil application of *Trichoderma asperellum* GDFS1009 granules promotes growth and resistance to *Fusarium graminearum* in maize. *J Integr Agric* [Internet]. 2019;18(3):599–606. Available from:  
<https://www.sciencedirect.com/science/article/pii/S2095311918620891>
35. Saravanakumar K, Li Y, Yu C, Wang QQ, Wang M, Sun J, et al. Effect of *Trichoderma harzianum* on maize rhizosphere microbiome and biocontrol of *Fusarium* Stalk rot. *Sci Rep* [Internet]. 2017;7(1):1–13. Doi: <http://dx.doi.org/10.1038/s41598-017-01680-w>
36. Elshahawy IE, El-Mohamedy RS. Biological control of *Pythium* damping-off and root-rot diseases of tomato using *Trichoderma* isolates employed alone or in combination. *J Plant Pathol* [Internet]. 2019;101(3):597–608. Available from: <https://link.springer.com/article/10.1007/s42161-019-00248-z>
37. Sánchez-Montesinos B, Diáñez F, Moreno-Gavira A, Gea FJ, Santos M. Plant growth promotion and biocontrol of *Pythium ultimum* by saline tolerant *trichoderma* isolates under salinity stress. *Int J Environ Res Public Health* [Internet]. 2019;16(11):1–11. Available from: Plant growth promotion and biocontrol of *Pythium ultimum* by saline tolerant *trichoderma* isolates under salinity stress
38. Tchameni SN, Cotârle M, Ghinea IO, Ampere M, Bedine B. Involvement of lytic enzymes and secondary metabolites produced by *Trichoderma* spp . in the biological control of *Pythium myriotylum*. *Int Microbiol* [Internet]. 2019; Available from: <https://link.springer.com/article/10.1007/s10123-019-00089-x>
39. Awad-Allah EFA, Shams AHM, Helaly AA, Ragheb EIM. Effective Applications of *Trichoderma* spp. as Biofertilizers and Biocontrol Agents Mitigate Tomato *Fusarium* Wilt Disease. *Agric* [Internet]. 2022;12(11). Available from: <https://www.mdpi.com/2077-0472/12/11/1950#:~:text=In conclusion%2C>

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

effective applications of, and development of cherry tomatoes.

40. Díaz-Gutiérrez C, Arroyave C, Llugany M, Poschenrieder C, Martos S, Peláez C. *Trichoderma asperellum* as a preventive and curative agent to control *Fusarium wilt* in *Stevia rebaudiana*. *Biol Control* [Internet]. 2021;155(September 2020). Available from: <https://www.sciencedirect.com/science/article/abs/pii/S1049964421000074>
41. Aoki Y, Haga S, Suzuki S. Direct antagonistic activity of chitinase produced by *Trichoderma* sp. SANA20 as biological control agent for grey mould caused by *Botrytis cinerea*. *Cogent Biol* [Internet]. 2020;6(1):1747903. Doi: <https://doi.org/10.1080/23312025.2020.1747903>
42. Risoli S, Cotrozzi L, Sarrocco S, Nuzzaci M, Pellegrini E, Vitti A. *Trichoderma*-Induced Resistance to *Botrytis cinerea* in *Solanum* Species: A Meta-Analysis. *Plants* [Internet]. 2022;11(2). Available from: <https://www.mdpi.com/2223-7747/11/2/180>
43. Wu X, Lyu Y, Ren H, Zhou F, Zhang X, Zhao X, et al. Degradation of oxalic acid by *Trichoderma afroharzianum* and its correlation with cell wall degrading enzymes in antagonizing *Botrytis cinerea*. *J Appl Microbiol*. 2022;133(5):2680–93.
44. Kouadri MEA, Bekkar AA, Zaim S. First report of using *Trichoderma longibrachiatum* as a biocontrol agent against *Macrophomina pseudophaseolina* causing charcoal rot disease of lentil in Algeria. *Egypt J Biol Pest Control* [Internet]. 2023;33(1). Doi: <https://doi.org/10.1186/s41938-023-00683-2>
45. Wang Z, Wang Z, Lu B, Quan X, Zhao G, Zhang Z, et al. Antagonistic potential of *Trichoderma* as a biocontrol agent against *Sclerotinia asari*. *Front Microbiol*. 2022;13(October):1–18.
46. Grano-Maldonado MI, Ramos-Payan R, Rivera-Chaparro F, Aguilar-Medina M, Romero-Quintana JG, Rodríguez-Santiago A, et al. First molecular characterization of *colletotrichum* sp. And *fusarium* sp. isolated from mangrove in Mexico and the antagonist effect of *trichoderma harzianum* as an effective biocontrol agent. *Plant Pathol J* [Internet]. 2021;37(5):465–75. Available from: <https://pubmed.ncbi.nlm.nih.gov/34847633/>

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

47. Yadav M, Dubey MK, Upadhyay RS. Systemic resistance in chilli pepper against anthracnose (Caused by *colletotrichum truncatum*) induced by *trichoderma harzianum*, *trichoderma asperellum* and *paenibacillus dendritiformis*. *J Fungi*. 2021;7(4).
48. Yadav M, Divyanshu K, Dubey MK, Rai A, Kumar S, Tripathi YN, et al. Plant growth promotion and differential expression of defense genes in chilli pepper against *Colletotrichum truncatum* induced by *Trichoderma asperellum* and *T. harzianum*. *BMC Microbiol* [Internet]. 2023;23(1):1–17. Doi: <https://doi.org/10.1186/s12866-023-02789-x>
49. Drahansky M, Paridah M., Moradbak A, Mohamed A., Owolabi F, Abdulwahab Taiwo, Asniza M, et al. We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists TOP 1%. Intech [Internet]. 2016;i(tourism):13. Available from: <https://www.intechopen.com/books/advanced-biometric-technologies/liveness-detection-in-biometrics>
50. Safari Motlagh MR, Abolghasemi M. The effect of *Trichoderma* spp. isolates on some morphological traits of canola inoculated with *Sclerotinia sclerotiorum* and evaluation of their efficacy in biological control of pathogen. *J Saudi Soc Agric Sci* [Internet]. 2022;21(4):217–31. Doi: <https://doi.org/10.1016/j.jssas.2021.08.004>
51. Silva LG, Camargo RC, Mascarin GM, Nunes PS de O, Dunlap C, Bettiol W. Dual functionality of *Trichoderma*: Biocontrol of *Sclerotinia sclerotiorum* and biostimulant of cotton plants. *Front Plant Sci* [Internet]. 2022;13(October):1–14. Available from: <https://www.frontiersin.org/articles/10.3389/fpls.2022.983127/full>
52. Mohamed BFF, Sallam NMA, Alamri SAM, Abo-Elyousr KAM, Mostafa YS, Hashem M. Approving the biocontrol method of potato wilt caused by *Ralstonia solanacearum* (Smith) using *Enterobacter cloacae* PS14 and *Trichoderma asperellum* T34. *Egypt J Biol Pest Control* [Internet]. 2020;30(1). Available from: <https://ejbpc.springeropen.com/articles/10.1186/s41938-020-00262-9>
53. Yan L, Khan RAA. Biological control of bacterial wilt in tomato through the metabolites produced by the biocontrol fungus, *Trichoderma harzianum*. *Egypt J*

Biol Pest Control [Internet]. 2021;31(1). Available from: <https://ejbpc.springeropen.com/articles/10.1186/s41938-020-00351-9>

54. Chien YC, Huang CH. Biocontrol of bacterial spot on tomato by foliar spray and growth medium application of *Bacillus amyloliquefaciens* and *Trichoderma asperellum*. *Eur J Plant Pathol* [Internet]. 2020;156(4):995–1003. Available from: <https://link.springer.com/article/10.1007/s10658-020-01947-5>
55. Saksirirat W, Chareerak P, Bunyatrachata W. Induced systemic resistance of biocontrol fungus, *Trichoderma* spp. against bacterial and gray leaf spot in tomatoes. *As J Food Ag-Ind* [Internet]. 2009;9:99–104. Available from: [https://www.researchgate.net/publication/308374894\\_Induced\\_systemic\\_resistance\\_of\\_biocontrol\\_fungus\\_Trichoderma\\_spp\\_against\\_bacterial\\_and\\_gray\\_leaf\\_spot\\_in\\_tomatoes](https://www.researchgate.net/publication/308374894_Induced_systemic_resistance_of_biocontrol_fungus_Trichoderma_spp_against_bacterial_and_gray_leaf_spot_in_tomatoes)
56. Hemeda NF, Deeb E. Evaluation of biological Control potential for different *Trichoderma* strains against Root-Knot Nematode *Meloidogyne javanica*. *J Adv Lab Res Biol E-ISSN* [Internet]. 2019;10(1):16–22. Available from: <https://media.neliti.com/media/publications/279147-evaluation-of-biological-control-potenti-a5a101e2.pdf>
57. Lubian C, Kuhn OJ, Portz RL, Agustinha AM, ... Biological control of *Meloidogyne javanica* in bean plants by *Hohenbuehelia* spp. and *Trichoderma koningiopsis*. *Arq do Inst.* [Internet]. 2021;1–9. Available from: <https://www.scielo.br/j/aib/a/ZWghn3nmhjVV7sXmtjvxNqz/?format=pdf>
58. Mukhtar T, Tariq-Khan M, Aslam MN. Bioefficacy of *Trichoderma* Species Against Javanese Root-Knot Nematode, *Meloidogyne javanica*, in Green Gram. *Gesunde Pflanz* [Internet]. 2021;73(3):265–72. Available from: <https://link.springer.com/article/10.1007/s10343-021-00544-8>
59. Zhang S, Gan Y, Liu J, Zhou J, Xu B. Optimization of the Fermentation Media and Parameters for the Bio-control Potential of *Trichoderma longibrachiatum* T6 Against Nematodes. *Front Microbiol* [Internet]. 2020;11(September):1–12. Available from: <https://www.frontiersin.org/articles/10.3389/fmicb.2020.574601/full>
60. Ghosh SK, Pal S. Entomopathogenic potential of *Trichoderma longibrachiatum*

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco



and its comparative evaluation with malathion against the insect pest *Leucinodes orbonalis*. *Environ Monit Assess* [Internet]. 2016;188(1):1–7. Available from: <https://pubmed.ncbi.nlm.nih.gov/26676413/>

61. Rodríguez-González A, Casquero PA, Cardoza RE, Gutiérrez S. Effect of trichodiene synthase encoding gene expression in *Trichoderma* strains on their effectiveness in the control of *Acanthoscelides obtectus*. *J Stored Prod Res* [Internet]. 2019;83:275–80. Available from: <https://www.sciencedirect.com/science/article/abs/pii/S0022474X19301869>
62. Coppola M, Cascone P, Di Lelio I, Woo SL, Lorito M, Rao R, et al. *Trichoderma atroviride* p1 colonization of tomato plants enhances both direct and indirect defense barriers against insects. *Front Physiol* [Internet]. 2019;10(July):1–12. Available from: <https://www.frontiersin.org/articles/10.3389/fphys.2019.00813/full>
63. Yao X, Guo H, Zhang K, Zhao M, Ruan J, Chen J. *Trichoderma* and its role in biological control of plant fungal and nematode disease. *Front Microbiol* [Internet]. 2023;14(May):1–15. Available from: <https://www.frontiersin.org/articles/10.3389/fmicb.2023.1160551/full>
64. Poveda J, Eugui D. Combined use of *Trichoderma* and beneficial bacteria (mainly *Bacillus* and *Pseudomonas*): Development of microbial synergistic bio-inoculants in sustainable agriculture. *Biol Control* [Internet]. 2022;176(November):105100. Doi: <https://doi.org/10.1016/j.biocontrol.2022.105100>
65. Hyde KD, Xu J, Rapior S, Jeewon R, Lumyong S, Niego AGT, et al. The amazing potential of fungi: 50 ways we can exploit fungi industrially. *Fungal Divers* [Internet]. 2019;97(1):1–136. Doi: <https://doi.org/10.1007/s13225-019-00430-9>
66. Dutta P, Mahanta M, Singh SB, Thakuria D, Deb L, Kumari A, et al. Molecular interaction between plants and *Trichoderma* species against soil-borne plant pathogens. *Front Plant Sci* [Internet]. 2023;14(May):1–22. Available from: <https://www.frontiersin.org/articles/10.3389/fpls.2023.1145715/full>
67. Tan B, Li Y, Deng D, Pan H, Zeng Y, Tan X, et al. Rhizosphere inoculation of *Nicotiana benthamiana* with *Trichoderma harzianum* TRA1-16 in controlled environment agriculture: Effects of varying light intensities on the mutualism-

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

parasitism interaction. *Front Plant Sci* [Internet]. 2022;13(October):1–15. Available from:

<https://www.frontiersin.org/articles/10.3389/fpls.2022.989155/full>

68. Etesami H, Jeong BR, Glick BR. Contribution of Arbuscular Mycorrhizal Fungi, Phosphate–Solubilizing Bacteria, and Silicon to P Uptake by Plant. *Front Plant Sci* [Internet]. 2021;12(July):1–29. Available from: Contribution of Arbuscular Mycorrhizal Fungi, Phosphate–Solubilizing Bacteria, and Silicon to P Uptake by Plant
69. Zehra A, Raytekar NA, Meena M, Swapnil P. Efficiency of microbial bio-agents as elicitors in plant defense mechanism under biotic stress: A review. *Curr Res Microb Sci* [Internet]. 2021;2:100054. Doi: <https://doi.org/10.1016/j.crmicr.2021.100054>
70. Ilham B, Noureddine C, Philippe G, Mohammed EG, Brahim E, Sophie A, et al. Induced systemic resistance (ISR) in arabidopsis thaliana by bacillus amyloliquefaciens and trichoderma harzianum used as seed treatments. *Agric* [Internet]. 2019;9(8). Available from: <https://hal.science/hal-02277415/>
71. Collinge DB, Jensen DF, Rabiey M, Sarrocco S, Shaw MW, Shaw RH. Biological control of plant diseases – What has been achieved and what is the direction? *Plant Pathol* [Internet]. 2022;71(5):1024–47. Available from: <https://bsppjournals.onlinelibrary.wiley.com/doi/10.1111/ppa.13555>
72. Martinez Y, Ribera J, Schwarze FWMR, De France K. Biotechnological development of Trichoderma-based formulations for biological control. *Appl Microbiol Biotechnol* [Internet]. 2023;107(18):5595–612. Doi: <https://doi.org/10.1007/s00253-023-12687-x>
73. Sood M, Kapoor D, Kumar V, Sheteiwiy MS. Trichoderma : The “ Secrets ” of a Multitalented. *Plants* [Internet]. 2020;9:762. Available from: <https://www.mdpi.com/2223-7747/9/6/762>
74. Fan H, Yao M, Wang H, Zhao D, Zhu X, Wang Y, et al. Isolation and effect of *Trichoderma citrinoviride* Snef1910 for the biological control of root-knot nematode, *Meloidogyne incognita*. *BMC Microbiol* [Internet]. 2020;20(1):1–11. Available from:

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

<https://bmcmicrobiol.biomedcentral.com/articles/10.1186/s12866-020-01984-4#:~:text=In conclusion%2C this study showed,masses and J2s on tomato.>

75. Mawar R, Manjunatha BL, Kumar S. Commercialization, Diffusion and Adoption of Bioformulations for Sustainable Disease Management in Indian Arid Agriculture: Prospects and Challenges. *Circ Econ Sustain* [Internet]. 2021;1(4):1367–85. Available from: <https://link.springer.com/article/10.1007/s43615-021-00089-y>
76. SENAVE. Registros de Productos [Internet]. Sitio Web. 2023 [cited 2023 May 5]. Available from: [http://secure.senave.gov.py:8443/registros/servlet/com.consultaregistros2.prod\\_agro2](http://secure.senave.gov.py:8443/registros/servlet/com.consultaregistros2.prod_agro2)
77. Ballagro Agro Tecnologia Ltda. ECOTRICH WP [Internet]. Ficha Técnica. San Pablo, Brasil: Ballagro Agro Tecnologia Ltda.; 2019.
78. Koppert Products. Trichodermil [Internet]. Ficha Técnica. 2023. Available from: <https://www.koppert.com.py/trichodermil/>
79. Simbiose. StimuControl [Internet]. Sitio Web. 2023 [cited 2023 May 5]. Available from: <https://simbiose-agro.com.br/product/view/stimucontrol#>
80. Rizobacter Argentina S.A. RIZODERMA [Internet]. Buenos Aires, Argentina: Rizobacter Argentina S.A.; 2018. p. 1–6. Available from: [https://www.syngenta.com.ar/sites/g/files/kgtney396/files/media/document/2020/04/21/marbete\\_rizoderma.pdf](https://www.syngenta.com.ar/sites/g/files/kgtney396/files/media/document/2020/04/21/marbete_rizoderma.pdf)
81. Lallemand Plant Care. LALSTOP QUALITY WG [Internet]. Sitio Web. 2023. Available from: <https://www.lallemandplantcare.com/en/usa/products/product-details/lalstop-g46-wg/>
82. Agro Advance Technology S.A. HULKGREEN [Internet]. Buenos Aires (Argentina): Agro Advance Technology S.A.; 2023. Available from: <https://www.agrofy.com.ar/biocontrolador-hulkgreenr.html>
83. Aqua Free. AQ TRC [Internet]. Nueva Italia, Paraguay: Aqua Free; 2023. p. 1. Available from: <https://www.af.com.py/productos/aq-trc>

Marchuk Larrea CN, Benítez Rodas GA, Sandoval-Espínola WJ, Arrúa PD, Lopez-Nicora H, Enciso-Maldonado G, et al. *Trichoderma* como agente biocontrolador- en foco

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